

EXOGENOUS CONTROL OF FOLLICULAR WAVE EMERGENCE IN CATTLE

G.A. Bo¹, G.P. Adams², R.A. Pierson³ and R.J. Mapletoft¹¹Departments of Herd Medicine and Theriogenology, and ²Veterinary Anatomy, W.C.V.M.³Department of Obstetrics and Gynecology, College of Medicine,
University of Saskatchewan, Saskatoon, Saskatchewan, Canada S7N 0W0.

ABSTRACT

Variability in ovarian response to superstimulatory treatments and in the interval from PGF_{2α} treatment to estrus in cattle is largely attributable to the status of follicular wave development at the time of treatment. To date, most treatments designed to control follicular wave development have been based on removal of the suppressive effect of the dominant follicle, either physically (by electrocauterization or ultrasound-guided follicle ablation) or hormonally (by GnRH or estradiol and progesterone treatment), and thereby induce the emergence of a new follicular wave at a specific time after treatment. Treatment of progesterone-implanted cattle with estradiol-17β (E-17β) resulted in suppression of the dominant follicle and emergence of a new follicular wave 4.3 ± 0.1 d later. Superstimulatory treatments initiated 4 d after E-17β treatment in progesterone-implanted cattle resulted in a superovulatory response comparable to that of cattle in which superstimulatory treatments were initiated on the second follicular wave. In another study, induced follicular wave emergence, regardless of the stage of the estrous cycle, resulted in similar superovulatory response and higher fertilization rates in heifers than when superstimulatory treatments were initiated 8 to 12 d after estrus (traditional approach). Finally, estrus synchronization treatments with E-17β plus progesterone and PGF_{2α} have resulted in synchronous estrus and ovulation. Overall, it appears that treatment with E-17β and progesterone in combination may be used to effectively control and synchronize follicular wave development and may have important implications in artificial control of ovarian cyclicity and superovulation.

INTRODUCTION

Ovarian asynchrony and variability in response to treatments remain the most limiting factors to the widespread implementation of advanced reproductive technologies in cattle, despite considerable progress in recent years (5,7,34). The status of follicular wave development is responsible for a large portion of the variability in ovarian response to superstimulatory

Key words: bovine, follicular wave emergence, estradiol, superovulation, estrus synchronization

Acknowledgments

Research was supported by the Natural Sciences and Engineering Research Council of Canada, the Catholic University of Córdoba, Argentina and the University of Saskatchewan. We thank Sanofi Inc., Overland Park, KS, USA, for Syncro-Mate-B; Coopers Agropharm Inc., Ajax, ON, Canada for Estrumate; and Vetrepharm Canada Inc., London, ON, for Folltropin-V and CIDR-B. Special thanks to our colleagues of the Instituto de Reproducción Animal de Córdoba (IRAC), Argentina and the University of Saskatchewan for technical assistance.

treatments and in the interval from PGF_{2α} treatment to estrus (26,34). The advent of real-time ultrasonography has provided a means to determine follicular status prior to gonadotropin treatments. However, it is difficult to precisely determine the status of follicular development on the basis of a single ultrasonographic examination and it is often not practical to serially examine individual animals in a commercial setting. An alternative approach is to control the development of a follicular wave so that superstimulatory treatments may be initiated at the most favourable time, i.e. when the number of follicles capable of responding to exogenous gonadotropins is maximal. Similarly, estrus synchronization programs may be adapted so that induction of luteolysis is coincident with the presence of a growing dominant follicle. The purpose of this paper is to address theoretical and practical aspects of bovine follicular development with respect to artificial control of follicular wave emergence and its application in superovulation and estrus synchronization.

FOLLICULAR WAVE DYNAMICS IN CATTLE

Ovarian follicular development in cattle is a dynamic sequence of organized events which has been described as wave-like (29,38,43). A wave of follicular development has been defined as a synchronous development of a large number of follicles 4 to 5 mm in diameter, followed by selection and growth of the dominant follicle and suppression of the subordinates (20,21). Most bovine estrous cycles have 2 or 3 follicular waves. Wave emergence was detected, on average, on the day of ovulation (Day 0) and Day 10 for two-wave cycles and on Days 0, 9 and 16 for three-wave cycles (20,21). However, there was a great variation in the proportion of animals exhibiting 2 or 3 waves per cycle and in the day of wave emergence, particularly of Wave 2. The genetic and environmental factors which influence this variability have not been clearly defined.

The dominant follicle of a follicular wave becomes larger than the other follicles and, through the production of steroidal and non-steroidal substances, suppresses the development of subordinate follicles and prevents the emergence of the next follicular wave (4,27,28). It has been recently shown that the apparent time of selection of the dominant follicle was coincident with a significant decrease in FSH concentrations (3), whereas, the emergence of a follicular wave is preceded by an increase in FSH concentrations, 1 or 2 d earlier (2). Determination of ovarian factors which inhibit gonadotropin secretion or interfere with folliculogenesis is important in the development of new approaches to the exogenous control of follicular development.

SYNCHRONIZATION OF FOLLICULAR WAVE DEVELOPMENT

There are several possible ways to synchronize follicular development. Most investigations have focused on removal of the suppressive effect of the dominant follicle (physically or hormonally) to allow the emergence of a new follicular wave at a specific time after treatment. The basis of this theory was derived from studies in which cauterization of the dominant follicle or its suppression with steroid-free bovine follicular fluid during the growing phase was followed by a premature FSH release and emergence of a new follicular wave (2,27,28).

Follicle Ablation

Studies in which the dominant follicle was electrocauterized showed that removal of the dominant follicle during its growing phase hastened the emergence of the next follicular wave (4,28). Based on these observations, a study was designed to test the hypothesis that ultrasound-guided follicle aspiration (used for oocyte retrieval in IVF programs), as a method of follicle ablation, would induce the synchronous emergence of a new follicular wave and synchronous ovulation after treatment with $\text{PGF}_{2\alpha}$ in heifers (9). Follicle ablation consisted of aspiration of all follicles ≥ 5 mm in diameter in heifers at unknown stages of the estrous cycle; all heifers (ablation and control group) received $\text{PGF}_{2\alpha}$ 4 d later. Daily ultrasonography revealed that follicle ablation resulted in a synchronous emergence of a new follicular wave 1.5 d later, and tightly synchronized ovulation after $\text{PGF}_{2\alpha}$ treatment. Ultrasound-guided transvaginal follicle ablation may be a useful method of manipulating follicular wave dynamics for estrus synchronization or oocyte retrieval and has been shown to be useful for superstimulation (17).

Hormonal Treatments

The ablation procedure was directed at removing the suppressive effect of the dominant follicle on the emergence of a new wave. Hormonal approaches have been directed at causing luteinization or atresia of the follicles present at the time of treatment. This has been accomplished by using hCG (40) or GnRH analogs (31,40) to induce follicle luteinization or ovulation, or by progestogens and estradiol to cause atresia of the dominant follicle (12).

Results of recent ultrasonographic and histologic studies have shown that treatment with a GnRH analog causes antral follicles to undergo atresia or induces ovulation and subsequent formation of a new CL (47). A new dominant follicle was identified ultrasonographically (by retrospective analysis) within 3 to 4 d after GnRH treatment and this follicle became the ovulatory follicle after $\text{PGF}_{2\alpha}$ -induced luteolysis (6 d after GnRH treatment; 47). In another study, GnRH treatment followed by prostaglandin 7 d later increased the number of heifers showing estrus within a 5-d period, and enhanced precision of synchrony 2 to 3 d after $\text{PGF}_{2\alpha}$ compared to heifers receiving $\text{PGF}_{2\alpha}$ alone (45). A 6- or 7-d interval between GnRH and $\text{PGF}_{2\alpha}$ resulted in a high degree of estrus synchronization with pregnancy rates comparable to those after treatment with 2 injections of $\text{PGF}_{2\alpha}$ 11 d apart (23,45,47).

Progestogen and Estrogen Treatments

Steroid hormones have also been used to alter follicle growth. Exogenous progesterone has been shown to suppress follicle growth in a dose-dependent manner (1,18,44) and estradiol has been shown to induce follicle atresia (19). The combined effects of estradiol and progestogens on follicular wave dynamics in cattle was inferred from preliminary studies designed to investigate the use of estradiol valerate (EV) and a progestogen ear implant^a in a superstimulatory regimen in beef cows (10). Ultrasonographic examinations revealed that EV treatment at the time of implant insertion (2 d after estrus) was associated with a reduction in the mean diameter of the largest and second largest follicles over a 5-d period followed by an increase in follicle

^a Syncro-Mate-B, Sanofi Inc., Overland Park., KS, U.S.A.

diameters, presumably from a new follicular wave. In a subsequent study (11), 5 mg EV given during the early growing phase (Day 1; ovulation=Day 0) suppressed the dominant follicle of the first follicular wave and resulted in early emergence of the next follicular wave in heifers without progesterone implants. However, treatment at the mid- (Day 3) or late-(Day 6) growing phase resulted in a delayed emergence of the next wave. This was attributed to an incomplete suppression of the dominant follicle combined with a prolonged effect of the valerate form of estradiol.

Because of the potential adverse effects of the prolonged action of EV, a series of experiments were designed to evaluate the effects of a shorter acting estrogen, estradiol-17 β (E-17 β). The first experiment was designed to test the hypothesis that E-17 β more effectively suppresses the dominant follicle when administered in combination with a progesterone ear implant than when given alone (12). In heifers treated with E-17 β plus progesterone ear implants, the dominant follicle ceased to grow 1 d after E-17 β treatment and subsequently regressed, resulting in an early emergence of the next follicular wave (Day 5.2 \pm 0.2). Conversely, E-17 β administration to heifers without progesterone implants did not effectively suppress the dominant follicle and emergence of the next wave was delayed (Day 9.8 \pm 1.1). In this study, the effect of E-17 β and progesterone on gonadotropins also was evaluated and results clearly showed that estradiol-induced LH release was not associated with follicle suppression. A post-treatment LH surge was detected in 5 of 6 heifers treated with E-17 β alone, and the dominant follicle was not suppressed, whereas heifers with progesterone implants did not exhibit a LH surge after E-17 β treatment and the dominant follicle regressed. Furthermore, in the heifers with progesterone implants plasma FSH concentrations decreased by 6 h after E-17 β treatment and increased gradually over a period of 24 to 42 h, whereas in heifers treated with E-17 β alone FSH decreased and then increased dramatically 12 h later, in association with the LH surge. It was concluded that E-17 β was more effective in inducing follicle suppression when combined with a progesterone ear implant. Also, this study provided the rationale for the hypothesis that the suppressive effect of E-17 β and progesterone in combination was due to suppression of both FSH and LH secretion, and that gonadotropin suppression must persist for at least 24 h to elicit complete follicle regression. In a second experiment designed to determine an effective dosage regimen of E-17 β for suppression of follicular growth in progesterone-implanted heifers, a single dose of 5 mg E-17 β was as effective as higher or repeated doses in inducing follicular suppression and resulted in consistent emergence of a new follicular wave 3 to 5 d later (12).

A third study evaluated the efficacy of E-17 β and progesterone treatment in synchronizing wave emergence when treatments were given at different stages of dominant follicle development (13). Beef cows and heifers were allocated into 4 treatment groups: untreated control animals and those that were given progesterone implants on Day 2, 5 or 8 and injected with 5 mg E-17 β im on Day 3, 6 or 9, respectively. Treatment days were expected to coincide with mid-growing phase (Day 3), late growing/early static phase (Day 6) and late static phase (Day 9) of the dominant follicle of the first wave (21). Day 9 was also expected to coincide with the emergence of the second follicular wave (12). The hypothesis that E-17 β plus progesterone treatment results in synchronous emergence of a new follicular wave was supported. Estradiol-17 β treatment resulted in an early emergence of the second follicular wave in heifers treated on Day 3 (Day 7.3 \pm 0.3) and delayed emergence in those treated on Day 6 (Day 10.7 \pm 0.2) compared to the control

group (Day 8.6 ±0.3; P<0.05). Additionally, treatment on Day 9 hastened emergence of the third wave (Day 13.1 ±0.3 vs Day 15.3 ±0.6 for Day 9 and control groups, respectively). The net result was that the interval from treatment to wave emergence was not different among E-17β groups and occurred, on average, 4.3 days after E-17β treatment. These results were consistent with those of the previous experiments (11) and are summarized in Table 1. Combined among experiments, the mean interval from E-17β treatment to follicular wave emergence in 47 animals was 4.3 ±0.1 d, and occurred 3 to 5 d after E-17β treatment in 44 of 47 (94%) heifers and cows treated. We concluded that treatment with E-17β and progestogen in combination can be used effectively to control and synchronize follicular wave emergence and may have important implications in artificial control of ovarian cyclicity and superovulation.

Table 1. Interval from estradiol-17β treatment to the emergence of the next follicular wave in heifers and cows with a progestogen ear implant (3 experiments combined; 11,12).

	5 mg Estradiol-17β			
	Day 1	Day 3	Day 6	Day 9
n	13	15	9	9
Wave Emergence (day)	4.5 ±0.2	4.0 ±0.3	4.6 ±0.2	4.1 ±0.3
(range)	(4 to 5)	(3 to 5)	(4 to 6)	(3 to 5)

Means among groups did not differ.

SYNCHRONIZED FOLLICULAR WAVE EMERGENCE AND SUPERSTIMULATION

Although there have been many reports on dosage regimens and types of gonadotropin preparations for ovarian superstimulation (34), it has been proposed that most of the variability in ovarian response to superstimulatory treatments in cattle is associated with variations in the status of follicular development at the time of treatment (5,7,34,35). In this regard, fewer ovulations were reported when superstimulatory treatments were initiated in the presence of a dominant follicle (17,22,25) or after selection of the dominant follicle (3,39). A higher superovulatory response has been reported when large numbers of small follicles were present at the time superstimulatory treatments were initiated (41). Two recent studies evaluated the responsiveness to superstimulatory treatments initiated with specific regard to follicular wave emergence (6,36). Results demonstrated that treatments initiated on the day before or the day of wave emergence resulted in a higher ovulatory response than treatments initiated 1 or 2 d later. Collectively, results clearly indicate that superstimulatory treatments must be initiated at the time of the endogenous pre-wave FSH surge or wave emergence (before dominant follicle selection) to obtain maximum superovulatory response in a given animal.

Two experiments were designed to determine the superovulatory response following artificially induced follicular wave emergence in cattle. The first experiment tested the hypothesis that E-17β plus progestogen treatment will induce a synchronized crop of follicles as responsive to exogenous gonadotropins as those of the second (spontaneous) follicular wave of the cycle. Beef cows either received progestogen ear implants on Day 0 (ovulation) plus 5 mg of E-17β im on Day 1 and were superstimulated on Day 5 or were not implanted (control) and superstimulated

on Day 8 (expected emergence of the second follicular wave; 11). Superstimulatory treatments consisted of 400 mg NIH-FSH-P1 of Folltropin-V^b administered by either a single sc injection behind the shoulder or divided into twice daily im injections over 4 d. Forty-eight hours after superstimulatory treatments were initiated, 500 µg cloprostenol^c was injected im and implants were removed 12 h later. Animals were inseminated 60 and 72 h after cloprostenol and slaughtered 7 d later for ova/embryo collection and CL counts. Results confirmed the hypothesis that E-17β treatment of progestogen-implanted cattle will induce a new wave of follicles as responsive to gonadotropin treatments as those of the second follicular wave (Table 2).

Table 2. Response (mean ±SEM) of control cows superstimulated with Folltropin-V on Day 8 after ovulation (Day 0) or cows that were given a progestogen ear implant on Day 0, 5 mg estradiol-17β (E-17β) on Day 1 and superstimulated on Day 5.

	Control		Progestogen plus E-17β	
	Single sc injection	Twice daily im injections	Single sc injection	Twice daily im injections
n	18	16	19	18
CL	22.0 ±3.5	23.7 ±3.7	27.0 ±3.1	16.6 ±3.4
Total ova/embryos	14.2 ±2.5	14.1 ±2.7	13.2 ±1.4	10.6 ±1.8
Fertilized ova	9.2 ±2.0	8.7 ±1.7	9.1 ±1.2	7.6 ±1.7
% Fertilized ova	65	62	69	72
Transferable embryos	5.4 ±1.5	5.5 ±1.7	5.4 ±1.0	3.9 ±1.5
% Transferable embryos	37	39	41	37

Means and percentages were not different

A second experiment was designed to compare superovulatory response following induced synchronized follicle growth by hormonal or physical treatments with a control group of beef heifers. The control group represented the traditional approach with heifers superstimulated between 8 and 12 d after observed estrus. Synchronized wave emergence was induced at unknown stages of the estrous cycle by a progestogen implant plus 5 mg of E-17β 1 d later (progestogen plus E-17β) or by ablating all follicles ≥5 mm in diameter using ultrasound-guided transvaginal follicle aspiration. An additional group of heifers treated with a progestogen ear implant alone also was included (progestogen alone). Superstimulation of the treated groups was initiated 5 d after progestogen implant insertion (4 d after E-17β) or 1.5 d after follicle ablation. Superstimulatory treatments consisted of 400 mg NIH-FSH-P1 Folltropin-V given by a single sc injection behind the shoulder. Cloprostenol treatments, implant removal, AI and ova/embryo collections were done as in Experiment 1. There were no differences among groups in the number of CL and total number of ova/embryos collected (Table 3). However, treatment with progestogen plus E-17β or follicle ablation resulted in a higher fertilization rate than the other treatment groups. Furthermore, the percentage of transferable embryos was higher in heifers treated with progestogen plus E-17β than those treated with progestogen alone.

^b Vetrepharm Canada Inc., London, ON, Canada.

^c Estrumate, Coopers Agropharm Inc., Ajax, ON, Canada.

Table 3. Response (mean \pm SEM) of beef heifers superstimulated between 8 and 12 d after estrus (control) or following induced synchronized follicle growth with progestogen plus estradiol-17 β (E-17 β) or follicle ablation at unknown stages of the estrous cycle.

	Control	Progestogen plus E-17 β	Progestogen alone	Ablation
n	18	19	19	20
CL	25.4 \pm 5.3	28.8 \pm 3.5	24.3 \pm 3.5	17.2 \pm 2.4
Total ova/embryos	8.9 \pm 1.5	13.2 \pm 2.1	12.4 \pm 2.2	7.8 \pm 1.3
Fertilized ova	6.0 \pm 1.1 ^a	11.2 \pm 2.0 ^b	7.5 \pm 1.9 ^{ab}	6.6 \pm 1.2
% Fertilized ova	68 ^a ^x	84 ^b	60 ^a	85 ^y
Transferable embryos	3.9 \pm 0.8	6.6 \pm 1.8	4.4 \pm 1.6	3.8 \pm 0.8
% Transferable embryos	44 ^{ab}	50 ^b	35 ^a	49

^{ab} Means and percentages (progestogen and control groups) with superscripts not in common are different ($P < 0.05$).

^{xy} Percentages of fertilized ova (ablation versus control group) are different ($P < 0.05$).

A recent study has shown that superovulatory response and especially the quality of embryos collected in heifers with progesterone releasing intravaginal devices (PRID) was influenced by the stage of the estrous cycle at the time of PRID insertion (24). The poor embryo quality in the group of heifers in which PRID were inserted in the early luteal phase was attributed to follicles that were under the influence of the dominant follicle and oocytes that were undergoing degeneration at the time superstimulatory treatments were initiated. Since heifers were implanted with progestogen at random stages of the estrous cycle in the present experiment, it is conceivable that the poor embryo quality in this group was a consequence of having follicles at various developmental stages at the time of superstimulation. In contrast, E-17 β treatment induced a new wave of follicles 4 d later and consequently a more uniform group of viable follicles was present at the time of superstimulation. Results from the present study demonstrated that induced follicular wave emergence, regardless of the stage of the estrous cycle, resulted in a superovulatory response at least comparable to the more traditional method of superstimulation 8 to 12 d post-estrus. These approaches obviate the necessity of detecting estrus prior to superstimulation treatments in donor cattle and may in fact result in more viable gonadotropin-responsive follicles.

ESTRUS SYNCHRONIZATION WITH PROGESTOGEN AND ESTRADIOL TREATMENTS

In estrus synchronization programs, the interval from PGF_{2 α} treatment to expression of estrus is determined by the stage of development of the dominant follicle at the time of treatment (26,42), which in turn affects pregnancy rates following fixed-time AI (30). Heifers with a viable dominant follicle returned to estrus in 48 to 60 h after PGF_{2 α} , whereas, heifers exhibited estrus in 5 to 7 d when the dominant follicle had started to undergo atresia. This prolonged interval is a reflection of the time required for a follicle from the new wave to grow and develop to a preovulatory state (26). Various progestogen/progesterone delivery systems and protocols have been developed for estrus synchronization (32,37); however, fertility was usually lower than in

untreated cattle when progestogen treatments lasted more than 14 d (32). Alternate, shorter progestogen treatment protocols (7 to 10 d) with $\text{PGF}_{2\alpha}$ given before or at the time of termination of treatments have been devised to improve fertility (32,37). However, this has not resulted in sufficient synchrony of estrus/ovulation for fixed-time AI. In addition, pregnancy rates were low when treatments were initiated during the late luteal phase (after Day 14; 8,16). Poor fertility after long-term progestogen treatments or short-term treatments initiated late in the estrous cycle was attributed to prolonged maintenance of the dominant follicle and ovulation of an aged oocyte (8,44). These results point out the need to synchronize follicular development in order to ensure the presence of a viable growing dominant follicle at the time of progesterone withdrawal and/or $\text{PGF}_{2\alpha}$ treatment (39).

A series of experiments were designed to determine whether E-17 β treatment of heifers with an intravaginal progesterone releasing device (CIDR-B) will reduce variation in the interval from treatment to ovulation, compared to more traditional methods of estrus synchronization in cattle. In a preliminary experiment involving 34 beef heifers (14), treatment with CIDR-B for 7 d plus 100 mg progesterone and 5 mg E-17 β im at the time of CIDR-B insertion and 500 μg cloprostenol at the time of CIDR-B removal, resulted in 75% of the heifers ovulating (detected by ultrasonography) between 72 and 84 h after CIDR-B removal. Only 40% of the heifers treated with 2 injections of $\text{PGF}_{2\alpha}$ 11 d apart, and 33% of heifers treated with CIDR-B without E-17 β ovulated during the same period of time ($P < 0.05$). Similar results were obtained in 2 other experiments in beef heifers treated with CIDR-B for 10 d and E-17 β administered 1 d after CIDR insertion or Brahman cows treated with CIDR-B for 10 d and E-17 β given 2 d after implant insertion (46). A higher degree of synchrony has also been shown when estradiol benzoate was used in combination with CIDR-B for 9 or 10 d (33). In yet another study^d, treatment with E-17 β and progesterone on the second day of an 8-d melengestrol acetate (MGA) treatment protocol (with $\text{PGF}_{2\alpha}$ on the last day) has resulted in improved pregnancy rates compared to MGA alone. Overall, results of these studies suggest that estradiol treatment in combination with progestogen/progesterone will cause the synchronous development of an ovulatory follicle and may be more efficacious than traditional approaches in synchronization of estrus and ovulation for timed-breeding.

SUMMARY

Imprecision in the degree of ovarian synchrony and variability in response to treatments have continued to be the most limiting factors in the application of new reproductive technologies in cattle. There is sufficient evidence to indicate that superstimulatory treatments must be initiated at the time of follicular wave emergence (spontaneous or induced) to obtain maximum superovulatory response. Synchrony of wave emergence can be accomplished by different approaches such as ultrasound-guided follicle ablation (wave emergence in 1.5 d), or treatments with estradiol and progestogen (wave emergence in 4 d) or possibly GnRH. Superstimulatory treatments initiated at the expected time of the induced wave emergence results in a superovulatory response at least comparable to treatments initiated on a spontaneous wave or to those initiated between 8 to 12 d after estrus. Synchronizing wave emergence in a group of randomly cycling animals obviates the need of estrus synchronization prior to superstimulation

^d JP Kastelic, unpublished.

and facilitates the management of embryo donors in large scale embryo transfer programs. Finally, incorporation of treatments that synchronize follicular wave emergence in estrus synchronization programs would insure the presence of a growing dominant follicle at the time of termination of progestogen treatments and/or PGF_{2α} treatment and result in synchronous estrus and ovulation, allowing for the effective use of fixed-time AI with high pregnancy rates.

REFERENCES

1. Adams GP, Matteri RL, Ginther OJ. Effect of progesterone on growth of ovarian follicles, emergence of follicular waves and circulating FSH in heifers. *J Reprod Fertil* 1992; 95:627-640.
2. Adams GP, Matteri RL, Kastelic JP, Ko JCH, Ginther OJ. Association between surges of follicle stimulating hormone and the emergence of follicular waves in heifers. *J Reprod Fertil* 1992; 94:177-188.
3. Adams GP, Kot K, Smith CA, Ginther OJ. Selection of a dominant follicle and suppression of follicular growth in heifers. *Anim Reprod Sci* 1993; 30:259-271.
4. Adams GP, Kot K, Smith CA, Ginther OJ. Effect of the dominant follicle on regression of its subordinates. *Can J Anim Sci* 1993; 73:267-275.
5. Adams GP. Control of ovarian follicular wave dynamics in cattle: implications for synchronization and superstimulation. *Theriogenology*, 1994; 41:19-24.
6. Adams GP, Nasser LF, Bo GA, Garcia A, Del Campo MR, Mapletoft RJ. Superstimulatory response of ovarian follicles of Wave 1 versus Wave 2 in heifers. *Theriogenology* 1994; 42:1103-1113.
7. Armstrong DT. Recent advances in superovulation of cattle. *Theriogenology* 1993; 39:7-24.
8. Beal WE, Chenault JR, Day ML, Corah LR. Variation in conception rates following synchronization of estrus with melengestrol acetate and prostaglandin F_{2α}. *J Anim Sci*. 1988; 66:599-602.
9. Bergfelt DR, Lightfoot KC, Adams GP. Ovarian synchronization following ultrasound-guided transvaginal follicle ablation in heifers. *Theriogenology* 1994; 42:895-907.
10. Bo GA, Pierson RA, Mapletoft RJ. The effect of estradiol valerate on follicular dynamics and superovulatory response in cows with Syncro-Mate-B implants. *Theriogenology* 1991; 36:169-183.
11. Bo GA, Nasser LF, Adams GP, Pierson RA, Mapletoft RJ. Effect of estradiol valerate on ovarian follicles, emergence of follicular waves and circulating gonadotropins in heifers. *Theriogenology* 1993; 40:225-239.
12. Bo GA, Adams GP, Pierson RA, Caccia M, Tribulo H, Mapletoft RJ. Follicular wave dynamics after estradiol-17β treatment of heifers with or without a progestogen implant. *Theriogenology* 1994; 41:1555-1569
13. Bo GA, Caccia M, Martinez M, Adams GP, Pierson RA, Mapletoft RJ. The use of estradiol-17β and progestogen treatment for the control of follicular wave dynamics in beef cattle. *Theriogenology* 1994; 40:165
14. Bo GA, Caccia M, Tribulo H, Adams GP, Pierson RA, Mapletoft RJ. Synchronous ovulation in heifers treated with E-17β and CIDR-B vaginal devices. *Proc Can Society Anim Sci, Regina, SK, 1994; 284 abstr.*
15. Bolt DJ, Scott V, Kiracofe GH. Plasma LH and FSH after estradiol, norgestomet and GnRH treatment in ovariectomized beef heifers. *Anim Reprod Sci* 1990; 23:263-271.
16. Brink JT, Kiracofe GH. Effect of estrous cycle stage at Syncro-Mate-B treatment on conception and time to estrus in cattle. *Theriogenology* 1988; 29:513-518.
17. Bungarts L, Niemann H. Assessment of the presence of a dominant follicle and selection of dairy cows suitable for superovulation by a single ultrasound examination. *J Reprod Fertil* 1994; 101:583-591.
18. Burke CR, Mihm M, Macmillan KL and Roche JF. Some effects of prematurely elevated concentrations of progesterone on luteal and follicular characteristics during the estrous cycle in heifers. *Anim Reprod Sci* 1994; 35:27-39.
19. Diershke DJ, Chaffin CL, Hutz RJ. Role and site of estrogen action in follicular atresia. *Trends Endocrinol Metab* 1994; 5:215-219.
20. Ginther OJ, Kastelic JP, Knopf L. Temporal associations among ovarian events in cattle during estrous cycles with two and three follicular waves. *J Reprod Fertil* 1989; 87:223-230.
21. Ginther OJ, Kastelic JP, Knopf L. Composition and characteristics of follicular waves during the bovine estrous cycle. *Anim Reprod Sci* 1989; 20:187-200.
22. Guilbault LA, Grasso F, Lussier JG, Matton P, Rouillier P. Decreased superovulatory responses in heifers superovulated in presence of a dominant follicle. *J Reprod Fertil* 1991; 91:81-89.

23. Guilbault LA, Villeneuve P, Laverdiere P, Proulx J, Dufour JJ. Estrus synchronization in beef cattle using a potent GnRH analog (Buserelin) and cloprostenol J Anim Sci Suppl 1991; 69:419 abstr.
24. Goulding D, Williams DH, Roche JF, Boland MP. Effect of exogenous progesterone on superovulatory response in heifers inseminated with fresh or frozen semen. J Reprod Fertil 1994; 100:505-510.
25. Huhtinen M, Raino V, Aalto J, Bredbacka P. Increased ovarian responses in the absence of the dominant follicle in superovulated cows. Theriogenology 1992; 37:457-463.
26. Kastelic JP, Knopf L, Ginther OJ. Effect of day of prostaglandin F_{2α} treatment on selection and development of the ovulatory follicle in heifers. Anim Reprod Sci 1990; 23:169-180.
27. Kastelic JP, Ko JCH, Ginther OJ. Suppression of dominant and subordinate ovarian follicles by a proteinaceous fraction of follicular fluid in heifers. Theriogenology 1990; 34:499-509.
28. Ko JCH, Kastelic JP, Del Campo MR, Ginther OJ. Effects of a dominant follicle on ovarian follicular dynamics during the estrous cycle in heifers. J Reprod Fertil 1991; 91:511-519.
29. Knopf L, Kastelic JP, Schallenberger E, Ginther OJ. Ovarian follicular dynamics in heifers: test of two-wave hypothesis by ultrasonically monitoring individual follicles. Dom Anim Endocr 1989; 6:111-119.
30. Macmillan KL, Henderson HV. Analyses of the variation in the interval from an injection of prostaglandin F_{2α} to estrus as a method of studying patterns of follicle development during dioestrus in dairy cows. Anim Reprod Sci 1984; 6:245-254.
31. Macmillan KL, Thatcher WW. Effects of an agonist of gonadotropin releasing hormone on ovarian follicles in cattle. Biol Reprod 1991; 45:883-889.
32. Macmillan KL, Peterson AJ. A new intravaginal progesterone releasing device for cattle (CIDR-B) for estrus synchronization, increasing pregnancy rates and the treatment of post-partum anestrus. Anim Reprod Sci 1993; 33:1-25.
33. Macmillan KL, Taufa VK, Day AM. Combination treatments for synchronizing estrus in dairy heifers. Proc New Zealand Soc Anim Prod 1993; 53:267-270.
34. Mapletoft RJ, Pierson RA. Factors affecting superovulation in the cow: practical considerations. IETS Embryo Transfer Newsletter, 1993; 11:14-24.
35. Monniaux D, Chupin D, Saumande J. Superovulatory responses of cattle. Theriogenology 1983; 19:55-81.
36. Nasser LF, Adams GP, Bo GA, Mapletoft RJ. Ovarian superstimulatory response relative to follicular wave emergence in heifers. Theriogenology 1993; 40:713-724.
37. Odde KG. A review of synchronization of estrus in postpartum cattle. J Anim Sci 1990; 68:817-830.
38. Pierson RA, Ginther OJ. Ultrasonic imaging of the ovaries and uterus in cattle. Theriogenology 1988; 29:21-37.
39. Pierson RA, Ginther OJ. Follicular populations during the estrous cycle in heifers III. Time of selection of the ovulatory follicle. Anim Reprod Sci 1988; 16:81-95.
40. Rajamahendran R, Sianangama PC. Effect of human chorionic gonadotropin (hCG) on dominant follicles in cows: accessory corpus luteum formation, progesterone production and pregnancy rates. J Reprod Fertil 1992; 95:577-584.
41. Romero A, Albert J, Brink Z, Seidel GE (Jr). Numbers of small follicles in ovaries affect superovulation response in cattle. Theriogenology 1991; 35:265 abstr.
42. Savio JD, Boland MP, Hynes N, Mattiacci MR, Roche JF. Will the first dominant follicle of the estrous cycle of heifers ovulate following luteolysis on Day 7? Theriogenology 1990; 33:677-687.
43. Savio JD, Keenan L, Boland MP, Roche JF. Pattern of growth of dominant follicles during the oestrous cycle in heifers. J Reprod Fertil 1988; 83:663-671.
44. Savio JD, Thatcher WW, Morns GR, Entwistle K, Drost M, Mattiacci, MR. 1993. Effects of induction of low plasma progesterone concentrations with a progesterone-releasing intravaginal device on follicular turnover and fertility in cattle. J Reprod Fertil 1993; 98:77-84.
45. Thatcher, WW, Drost M, Savio JD, MacMillan KL, Entwistle KW, Schmitt EJ, De La Sota RL. New clinical uses of GnRH and its analogues in cattle. Anim Reprod Sci 1993; 33:27-49.
46. Tribulo HE, Bo GA, Kastelic JP, Pawlyshyn V., Barth AD, Mapletoft RJ. Estrus synchronization in cattle with estradiol-17β and CIDR-B vaginal devices. Theriogenology 1995, 43: In press-abstr.
47. Twagiramungu H, Guilbault LA, Proulx JG, Dufour J. Influence of corpus luteum and induced ovulation on ovarian follicular dynamics in postpartum cyclic cows treated with Buserelin and cloprostenol. J Anim Sci 1994; 42:1796-1805.